

Vol. 23, No. 3

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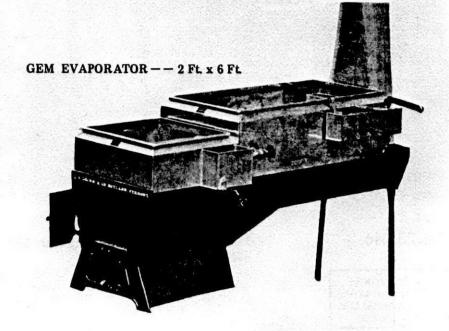
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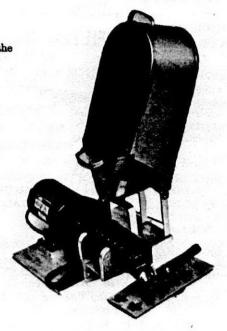
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THE MAPLE SYRUP

Published & Edited by:

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Editorial

First, the bad news: Maple Industry loses two friends.

A great loss occurred to the maple industry in early March when Aaron Hall was killed in an auto accident. Aaron was a part owner of Maple Syrup Supplies Co. of Potterville, Michigan and will be greatly missed by his friends and customers.

About the same time, Eugene Running, of Cambridge Springs, Pa. passed away after a long illness. Gene was well known and respected within the Pennsylvania and Ohio maple business and will surely be missed by all who knew him.

Now the good news:

It looks like the surplus of syrup is gradually diminishing. It's getting harder to find good syrup in bulk in New York State and I hear it's the same elsewhere except where there may have been a better crop. Some producers are hanging on to some waiting for a better price which is gradually creeping up.

There must be plenty left in Canada because of the amount coming across the border at prices we can't even compete with. But there's still plenty of market for top quality products. We just have to work harder to find them.

I don't have anything more really urgent to write about so I won't waste space. Hope you've all had as nice a summer as we have.

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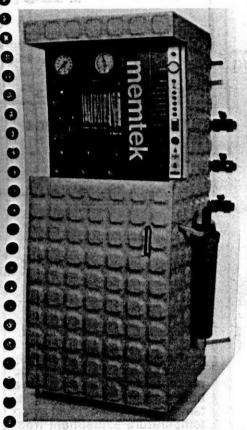
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FROM THE CHAIRMAN



Another summer has just about gone and the cooler weather has many producers thinking about sugaring again. The summer programs have been very well attended and enthusiasm is high. Several people even predicted a better than average crop next Spring. The favorite reason for a better crop is the large amount of sunshine we are having this summer. Many factors influence the sap run as well as the sugar content. Some day we may know the effect of dry weather, insect damage, acid rain and the many other conditions that



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cause stress in our maples.

The over-supply of syrup seems to be dwindling as more emphasis is being placed on marketing. I even hear of small increases in the price of bulk syrup. We have a very unique product and once the consumer becomes exposed to Pure Maple he seldom is satisfied with the blends or imitations. Any effort to inform the public about Pure Maple Products will in the long run help the entire industry.

The 24th annual meeting to be held in Traverse City, Michigan on October 24 and 25 is all set. Terry Healey and his committee have planned a great program for us.

Next year will be our 25th year and I hope it will be a banner year for the Council.

I am asking the directors to enter into a discussion concerning the **Maple Digest** the second day at Michigan. The Council will be 25 years old next year. We should reevaluate the **Digest** as it is the most important voice of the Council. I want discussion on:

- 1. Do you think the **Digest** does its job?
- 2. Suggestions for improvement.
- 3. Talk to advertizers and see if they feel they get their money's worth.
- 4. The **Digest** is the responsibility of the Council and reflects the work and enthusiasm of the members. It is up to the directors to insist that the **Digest** does the best job possible in representing the Council and the Maple Industry.

Please give this subject you concern.

Russ Davenport



Roger Sage presents Dr. Robert Morrow a plaque as Lew Staats looks on.

1983 New York Maple Tour

By Lewis J. Staats

The 1983 New York Maple Tour took place on August 1 and 2 in Essex and Franklin counties of Northern New York.

The tour was based at the Hotel Saranac of Paul Smith's College. Buses were used for transporting the group throughout the tour. The value of this means of transportation became obvious as the group traveled through the heavy tourist use villages and mountain roads.

On Monday, August 1, the groups journeyed to Lake Placid to tour the Olympic Ski Jump Complex and the Uihlein Sugar Maple Research-Extension Field Station. Information and demonstration stops included past research summaries, new research on the use of vapor compression

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evaporation, sap collection using tubing and vacuum systems, thinning and sugar bush management, insect detection and identification, testing individual tree sap sweetness with the use of a refractometer, and a tour and discussion of the clonal bank and progeny sugar maple plantations.

The banquet and program were held on the evening of August 1 at the Hotel Saranac. 260 tour participants were served an excellent roast beef dinner. Masters of ceremony were Guy Hutt, Cooperative Extension Agent in Essex County and Bill Pauling, Cooperative Extension Agent in Clinton County. First of the speakers was Dr. Harry Miller, acting president of Paul Smith's College. who welcomed the tour group to Hotel Saranac and gave an overview of Paul Smith's College. Several members of Cornell University staff spoke on Cornell's interest in maple syrup production including; Dr. Kenneth E. Wing, Associate Dean; Dr. Ray Oglesby. Chairman of the Department of Natural Resources: Dr. John Kelley, Department of Natural Resources; and Lewis Staats, Extension Specialist, Maple Syrup Production. Roger Sage, President of the New York State Maple Producers Association, discussed programs and activities of the Association. The reigning New York State Maple Queen, Miss Jill

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HAROLD TYLER MAPLE FARMS 607-638-9474 Westford, N.Y. 13488 Komrowski, was introduced. Dr. Robert Morrow, recently retired Professor of Forestry from Cornell University was presented a plaque from the New York State Producers Association in recognition for his contributions to the Maple syrup industry.

Dr. Morrow presented a history of the Uihlein-Cornell maple program emphasising the valuable support and gifts from Mr. and Mrs. Henry Uihlein of Lake Placid. Mr. and Mrs. Uihlein were then honored with a plaque presented by the New York State Maple Producers Association.

Tuesday morning, the tour traveled through the Adirondacks to visit Mt. Van Hovenberg bobsled and luge runs and three very nice maple operations in the Keene and Keene Valley area. The first maple stop was Frog Alley

Farm operated by Harold Heald in Keene Valley. Here syrup is produced on a 2.5 by 8 wood fired evaporator from 475 buckets on leased trees. Second stop found the group at Wade and Jud Whitney's family operation in Keene. This operation consists of a 5' by 14' wood fired evaporator with preheater which handles 2000 taps, 1600 of which are on vacuum. The last stop of the day was at the Alstead Hill Sugar House. This has a 5' by 15' wood fired evaporator and a sugar bush being improved by thinnings.

In finishing the 1983 New York Maple Tour, tour participants were given the opportunity to view equipment exhibits at the tour base in Saranac Lake from early afternoon until 5 pm.

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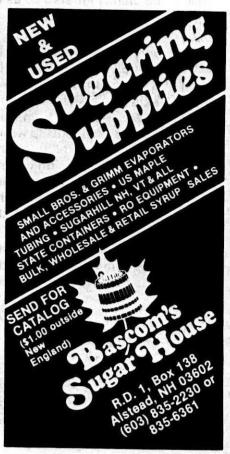
Mr. and Mrs. Henry Uihlein II Honored By The New York Maple Producers Association

By John W. Kelley Lewis J. Staats

At the 1983 New York Maple Tour banquet held August 1 at the Hotel Saranac of Paul Smith's College in Saranac Lake, the New York State Maple Producers Association presented Mr. and Mrs. Henry Uihlein II of Lake Placid a plaque in appreciation of their many years of generous support to maple syrup research.

The Uihleins have supported agricultural research at Cornell University for many years. In 1965, the Uihleins made the sugar bush portion of their Heaven Hill Farm at Lake Placid available to Cornell University for research and demonstration projects in maple syrup production. In addition to the lease of these forest lands, the Uihleins provided numerous gifts to help develop facilities and to obtain equipment for the direct support of the Cornell maple program. In 1981 the Uihleins deeded to the University 200 acres of the sugar bush as a gift. Also included in this gift was an 8acre parcel of land to be used by Cornell in cooperation with the U.S. Forest Service for a genetics study of "sweet tree" sugar maples. Progeny and clonal seed tree bank plantations were established on this site in the spring of 1983.

The Uihlein Sugar Maple Research - Extension Field Station has been named in honored of Mr. and Mrs. Henry Uihlein II. Cornell University and the New York State Maple Producers Association recognize and appreciate the continued support and interest of the Uihleins in the furtherance of maple syrup research and extension.





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Report On The Raithby Economizer

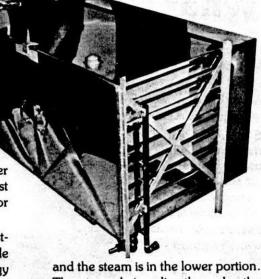
Subject: The Raithby Economizer placed at Mr. Earl Parker's of West Chazy, N.Y. by Leader Evaporator Co., Inc.

This unit is much like the unit tested by Bill Langenberg of Kemptville College of Agricultural Technology in Ontario.

The principle of the Economizer is to evaporate sap at temperatures below the boiling point.

Design of the Economizer:

The economizer consists of 8 - 4x8 stainless steel trays (Ontario unit had aluminum trays) providing 256 sq. ft. of surface area for evaporation. The travs are stacked one over the other and the sap is routed downward through each tray. A 24" tubeaxial fan is used to draw air over the sap in each tray which creates a negative pressure allowing the sap to steam at a lower temperature. This fan also removes the steam coming off the sap. A 1/2" gear pump driven by a 1/3 H.P. electric motor is used to pump the sap from the economizer to the evaporator. Steam produced by the main evaporator is the fuel used to operate the economizer. By the use of an air tight hood the steam is ducted from the evaporator to the economizer. The travs are double walled travs. The sap is in the upper portion of the tray



and the steam is in the lower portion.

The steam being directly under the sap heats it and causes it to evaporate.

We have made a few changes to the unit that was designed and tested in Ontario and found the results to be very favorable. The unit tested in Ontario showed a 65% increase in evaporation. The unit tested at Mr. Parker's, we feel showed a substantial increase over the previous years testing. Because of lack of time we were unable to install a flow meter but intend to do this next spring. We have based our results on sugar content only. 21/2% sap going into the economizer came out at 4.6%, 2% came out at 4.2%, 1.6% came out at 3.8%.

We feel the increase is the direct result of using a heat exchanger (preheater) and the routing of the steam.

We plan to do further testing of this unit at the start of the next sugar season.

Look for a future article for more promising results.

13

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Dear, Sir:

We sell maple syrup and candy and have been using your Kress jugs for eight years ..., we have a lot of comments about your jugs. Our customers love them. We are pleased with the quality of your jugs as we had used others before we used yours.

John & Carol Deets Titusville, PA Dear Sir

We have used Kress jugs purchased through M.M.S.A., so we know they are good. Our customers say they are "cute".

Paul Bergmann Traverse City. MI

Dear Kress Creations:

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Randy Whitcher Warren, NH

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THE SUGAR MAPLE BORER

Douglas C. Allen
Forest Entomologist, College of
Environmental Science and
Forestry Syracuse, New York

Lewis J. Staats
Extension Specialist, Uhlein-Cornell
Experimental Sugar Bush,
Lake Placid, New York

Few scenes distress a woodlot owner more than scarred tree trunks. This unsightly damage is of special concern to maple syrup producers. because a sugar maple's ability to produce sap is, for the most part, determined by tree vigor and general health. The sugar maple borer rarely kills trees, but it can be a major cause of trunk and crown damage in sugarbushes. In some regions, incidence of borer activity is low, but in many areas, 25 to 50 percent of the sugar maples may be attacked. Recognition of borer damage and an understanding of the ecological conditions that favor the insect will help woodlot owners reduce the economic impact of this pest.

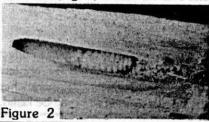
The adult sugar maple borer is a black beetle distinctly marked with bright yellow bands of varying width and shape (Fig. 1). The beetle is ap-



Figure 1.

proximately one inch long, and it belongs to a group commonly referred to as longhorn beetles, a name evoked by the unusually long antennae, or feeler-like structures, that are attached to the beetles' head.

The beetle deposits one to a few eggs in crevices or holes that it chews through the bark, usually on the basal 20 feet of a tree trunk. Many trees are probably used for egg laying, but the more vigorous ones are able to overcome the newly hatched larvae. Following egg hatch, the white, grublike larva (Fig. 2) enters the tree and



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Hardscrabble Tree Farm 35 Bridge Street Colebrook, NH 03576 feeds beneath the bark. Eventually it excavates a shallow transverse or oblique feeding gallery in the sapwood and inner bark (Fig. 3). As a result of



this girdling, which is similar in effect to the damage caused by an axe blaze or logging scar, large branches above the feeding site may be killed. This is important to sugarbush operators, because the amount of sap produced by a tree is primarily a function of crown size. Larval feeding also destroys inner bark tissues in a large area adjacent to the site where feeding actually takes place. A large catfaced scar or area of exposed wood is often produced (Fig. 3) and a significant portion of the tree trunk may be rendered unusable for tapping. Sugar maple borer scars are not always easy to detect. Sometimes they are conspicuous, but often the damage is hidden beneath slightly cracked and loosened bark (Fig. 4). The presence of the larval gallery engraved on the surface of the exposed sapwood (Fig. 3) distinguishes scars caused by sugar maple borer feeding from scars that are caused by other agents, such as fungi.

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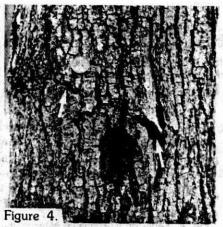
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The borer requires two years to complete development from egg to adult. In preparation for overwintering during the second year, the fully grown larva excavates an oval, 3/8-5/8 inch diameter gallery that penetrates the sapwood to a depth of 2-4 inches (Fig. 2). This may severely

degrade the first board that is sawn from that part of the log. In addition to this physical damage, borer attack stimulates a physiological response in sugar maple that protects the tree. but may further reduce the quality of lumber cut from previously infested sawlogs. For example, chemical barriers and callus tissue develop in the vicinity of borer injury. This is the tree's way of compartmentalizing the damage to prevent invasion of healthy tissue by wood-inhabiting microorganisms. The callus imparts a twisted grain to newly formed wood adjacent to the wound, while the chemical barrier results in a mineral stain, giving an undesirable color to lumber cut from the injured portion of the log.

Recently completed research at the State University of New York,



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College of Environmental Science and Forestry, showed that successful sugar maple borer attacks are generally limited to sugar maples that have been stressed and are in a weakened state. For the maple syrup producer. a program of proper sugarbush management that encourages maximum sap production, rapid growth and vigorous trees will help to reduce the incidence of damage. For example, proper thinning of stands during the highly susceptible pole timber stage, when tree diameters are between 5 to 11 inches, is especially important in a program of preventative maintenance. If sawtimber is the management objective, removal of previous-

ly damaged trees is also recommended to improve the quality of the residual stand and alot growing space to sound, and presumably, more valuable trees.

The sugar bush operator should use careful judgment, however, before condeming a tree. It is not necessary to remove sugar maples at the first sight of damage. As long as a tree pays its' way in terms of sap production it should be preserved.

Fig. 1. Sugar maple borer adult. Fig. 2. Larva of sugar maple borer in the overwintering cell.

Fig. 3. Damage caused by larval feeding. Arrow indicates transverse feeding gallery.

Fig. 4. Appearance of loosened and cracked bark that often conceals borer damage (arrows).



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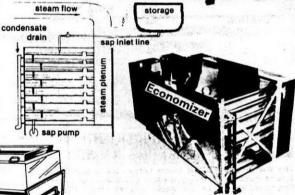


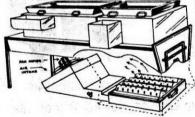
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where centers the sweetness he craves, a close investigation is attempted, during which his inquisitive nose is burned. This calls out a roar of pain and a stay in the proceedings. But after sundry whimperings and doleful head shakings, he rises to the occasion and looks in the tempting dish. The sight proves too much for his fortitude; down goes a big brown paw, and with a howl of rage is withdrawn and thrust in bruin's capacious mouth. This is repeated again and again, amid grunts of satisfaction and whines of pain. By this time the courage of the quartet has revived and there comes a rattling volley from the rifles, followed by a general stampede. The listeners in the cabin, not daring to venture out, can judge only by the growls and roars of agony, that some of their shots have taken effect. A search next morning discovers an enormous bear, quite dead, his glossy coat bedaubed with syrup, while his right paw, blistered and burned clean of hair, marked him as a native born New Englander in his love for maple syrup.

This article was written by Laura Sanderson and found in a history of Franklin County, Mass. *Picturesque Franklin 1891* by Russell M. Davenport.

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Microorganisms on Plastic Tubing Walls.

By Warren King¹ and Mariafranca Morselli

Sugar maple sap contains water and all the precursors of color and flavor, which develop when the sap is boiled to syrup. These precursors, mainly a sugar, sucrose, and small amounts of amino acids, phenolics. organic acids, and mineral irons, are subject to change during a sap flow season due to the metabolic changes in the tree, mediated mainly by temperature. Genetic differences among sugar maple trees also may play a role in these changes. But sugar maple sap also is a good growth medium for microorganisms. Depending on ambient temperature, microorganisms

act on the sap biochemistry at the taphole and in the sap collecting and storing systems.

The metabolic activity of microorganisms in sugar maple sap includes the enzymatic hydrolysis of some sucrose molecules, causing a small increase in sap invert sugar (mainly glucose) that contributes to the darkening of the syrup and to the development of the caramel flavor. Hence, microbial contaminants in the sap from tapholes, collecting systems, and storage tanks change syrup grade (1,2,3,4).

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Since the 1950's, because of research by Willits and colleagues, the maple syrup industry has recognized that sugarbush sanitation is of the highest priority for producing clean sap, and in turn, light-colored table syrup with good maple flavor. The profit of a sound maple sugaring operation depends largely on the quality of the syrup and the highest financial return is derived from the production of table grade syrups.

The widespread use of plastic tubing systems in sugaring operations has eliminated the extensive labor needed to collect sap from buckets. Furthermore, the use of vacuum pumps, which increase the rate of sap flow, has doubled sap yield. Sap quality problems arise if the tubing sags and if the sap lingers within the tubings or the large conduits. Maple producers have postulated that, even when the tubing system is properly positioned, the sap flow will decrease, because of microorganism build-up on the plastic walls, at the tubing connection with the spout, and in other critical spots.

Previous work has documented the number of bacteria and yeasts found in maple sap from different collecting and storing systems (2,3,4) but no research has been reported on the identification of microorganisms adhering to the plastic tubings.

With funding, in part, from the Chittenden County Sugar Makers Association, we have initiated a study to identify the microorganisms adhering to the internal walls of plastic tubings.2

In one experiment we compared the number and type of microor-

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ganisms found in early, mid-season, and late saps with the microorganisms found adhering to 5 feet of tubing used to collect the same saps. The sap was collected by gravity in a sterile flask from one taphole drilled with a bit and filled with a plastic spout, both rinsed in 10% bleach solution. The tubing from the spout was new. The tubing sections were cut and handled in sterile conditions.

For the experiment flushing the tubing with a sterile phosphate buffered saline solution and swabbing the tubing walls in 5 different areas proved to be more effective for removing microorganisms than did scraping the walls. The flushing technique was designed to remove any organism loosely attached to the tubing walls, while the swabbing technique was used to remove organisms strongly adhering to the walls. The microorganisms from the flushing solutions and swabs then were incubated for 3 days at 27°C (81°F) in the media proper for each type. Five biochemical tests then were performed to identify the organisms (5), and for each test, controls were run with known microorganism cultures (AT-CC3) These procedures will be published elsewhere.

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In a second experiment, sterilized sap and trypticase soy broth (a sterile nutrient solution) were inoculated into 25, 1-foot long, sections of the tubing used during the previous sap season at the Proctor Maple Research Farm. The tubings had been washed with a 10% bleach solution and rinsed with hot water immediately after the previous sap season and then were stored during summer and fall in a large dark, screened, and aerated area at the Farm. Cut. sealed (air. permeable) tubing sections containing the liquid were incubated for five intervals up to 2 weeks at 27°C (81°F) to allow for bacteria and yeasts, presumably present after storage in either a dry or spore form, a chance to grow. We flushed and swabbed the tubings at each interval, and then incubated the microorganisms in different nutrient media, and used the same identification procedures as in the first experiment (5).

A third experiment consisted in inoculating five new clean tubing sections with slightly contaminated sap. The same procedures as in the other experiments were used for incubation

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American Maple Museum P.O. Box 81 Croghan, N.Y. 13327 and identification of microorganisms.

Results and Conclusions:

The results of the first experiment are given in Table 1. Early in the sap season, tubings and sap were void of microorganisms, we believe because the taphole had been drilled with a clean procedure, although we did not use the aseptic procedure we use to collect sterile sap. However, a dramatic increase in microorganisms, although not at the same rate, was found in the flushing solutions at mid- and late-sap season in microorganisms adhering to the plastic tubings. Futhermore, even after the tubings were flushed, microorganisms still were found by swabbing the walls. More pseudomonads (fluorescent and nonfluorescent) from the tree bark and enterics from human contamination were recovered by flushing and swabbing procedures than were found in the sap. If we consider that flushing the tubing should dilute the microorganism sample, then we conclude that increase of microorganisms in flushed and swabbed samples means that adherence of microorganisms to the walls did indeed occur. Strong evidence also indicates that more adherence occurred in the tubing sections closer to the spout.

The results of the second experiment, designed to establish the number and type of microorganisms in the tubings stored for more than 6 months, surprisingly did not show any surviving microorganisms--not even one colony--for all the different techniques used. We hypothesize that the thorough bleach washing of the tubings preceding storage and/or high summer temperatures have kil-

led the microorganisms, both sporeand nonspore-forming organisms. If any spore had been present in the tubings, the length of the incubation period would have allowed them to germinate.

In the third experiment, designed to recover microorganisms deposited from sap on new tubing walls, about 20 microorganism types present in sap readily adhered to the tubing wall surface. This experiment will be repeated and confirmed. Microorganism types will be published elsewhere.

Recommendations:

More research is needed to confirm these preliminary findings from one season, one tree, one location and one type of tubing, and to enable us to extend specific recommendations how to clean tubing walls more effectively. However, in light of these findings, we recommend using the cleanest tapping technique possible in a sugarbush; also, washing the tubing and conduit systems at least once during the sap season and after it. The practice of washing the tubings only before the sap season starts,

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Table 1. Average microorganisms in sap and on tubing walls, reported in colony forming units per milliliter (CFU/ml).

					Sap Season	ason				
131			Early	EN	MSA MA AND MARTIN	PiW			Late	
Microorganisms			Tubing walls	walls	4 20	Tubing	Tubing walls		Tubing walls	walls
Types	3131	Sap	Flush	Swabbing	Sap	Flushing	Swabbing	Sap	Flushing	Swabbing
TOTAL	0 × 10	100	0 x 100	0 × 100	3.8x10 ²	2.1×10 ²	6.0x10 ¹	4.8x10 ⁵	1.8×10 ⁵	1.4×10 ³
TOTAL	0 × 1	100	0 x 100	0 × 100	8.1x10 ³	1.1x10 ⁴	5.0x10 ²	8.0x10 ⁵	l	2.8×10 ⁶
TOTAL AEROBES & FLUORESCENT PSEUDOMONADS	O X	100	0 × 10 ⁰	0 × 10 ⁰	8.9×10 ³	1.8×10 ⁴	5.4×10 ²	1.1×10 ⁶	2.0x10 ⁷	4.9×10 ⁴
TOTAL	0 × 1	100	0 × 10	0 x 100	4.0x10 ²	1.0x10 ³	2.1x10 ²	2.4×10 ²	6.0x10 ⁵	8.7×10 ²
TOTAL GRAM POSITIVES	0 × 1(100	0 × 10 ⁰	0 × 10 ⁰	1.1x10 ²	1.6×10 ²	0 ×10 ⁰	1.2×10 ⁵	1.2×10 ⁵	1.5×10 ³
TOTAL	0 × 1	100	0 × 100	0 x 100	4.4x10 ³	1.1×10 ⁵	2.2×10 ⁴	7.1x10 ⁵	2.2×10 ⁶	2.7×10 ⁴
TOTAL FERMENTERS. & NON-FERMENTERS	0 × 1	100	0 x 10 ⁰	0 × 10 ⁰	2.9×10 ²	La Contraction	1.0x10 ¹	1.0×10 ²	3.8×10 ⁵	7.0x10 ²

or just letting the first sap run out of the system, is not sufficient to maintain clean tubings, hence clean sap, throughout the season.

¹Undergraduate student and research associate professor, respectively, Botany Department, Maple Research Laboratory, Agricultural Experiment Station, University of Vermont.

periment Station, University of Vermont.

Lamb Tubing System. Robert M. Lamb,
Bernhards Bay, N.Y. Use of name of manufacturer does not imply endorsement by the
Vermont Agricultural Experiment Station over
others not named.

³American Type Culture Collection, Rockville, Md.

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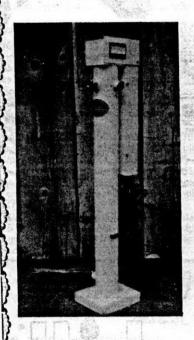




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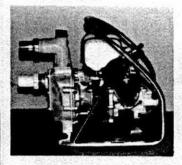
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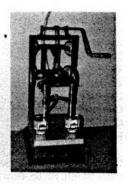
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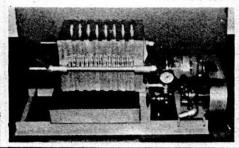
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Thoroughly tested by both producers and packers. Filtering capacity of up to 150 gallons.

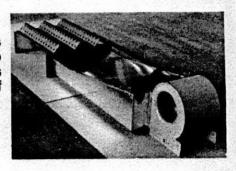
Have you checked on the Raithby Economiser? Maybe this will give your operation the needed increase in production. We are a licensed manufacturer for the U.S.A. Give us a call.

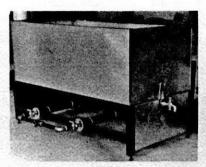
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FOR THE MAPLE SYRUP PRODUCER

The LEADER air induced units used with wood firing are set up with 4 speed direct drive blowers to offer the correct amount of forced air.

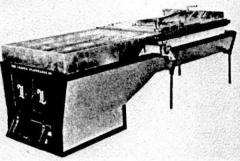
Use only dry, seasoned wood.





The LEADER 16" x 32" gas fired finish unit is ideal for the small canning operations and "boiling down" to make candy or sugar. 2 projected port ring burners rated at 33,000 BTUs for a well balanced unit.

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